Targeting the anti-inflammatory properties of the vagus nerve through vagus nerve stimulation: Therapeutic implications for inflammatory bowel diseases and other inflammatory conditions

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Abstract. The vagus nerve (VN) is the longest nerve of the organism that innervates major organs such as the heart, lungs and gastro-intestinal tract and is a major component of the autonomic nervous system at the interface of the central nervous system and the body. The VN has anti-inflammatory properties both through its afferent fibers, activating the hypothalamic-pituitary adrenal axis, and its efferent fibers activating the cholinergic anti-inflammatory pathway. Targeting these anti-inflammatory pathways is of interest for various inflammatory conditions. Bioelectronic medicine through VN stimulation (VNS) appears as an interesting tool to release inflammatory conditions. VNS is approved for the treatment of drug-resistant epilepsy and has potential therapeutic applications in chronic inflammatory disorders such as inflammatory bowel diseases and others. Preclinical data and pilot clinical studies argue for such an effect. However, larger randomized double-blinded control study and a long-lasting follow-up of the patients to confirm these promising results are awaiting. In addition, the optimal neurostimulation parameters to better treat common conditions and diseases that involve immune regulation need to be determined.

Keywords: vagus nerve, vagus nerve stimulation, cholinergic anti-inflammatory pathway, hypothalamic-pituitary adrenal axis, inflammatory bowel diseases, rheumatoid arthritis.

Introduction

The vagus nerve (VN), the tenth cranial nerve, is the longest nerve of the organism, which innervates major organs, such as the heart, lungs, and gastrointestinal tract. The vagus nerves are normally referred to in the singular. The VN is a major component of the parasympathetic nervous system, which, together with the sympathetic nervous system, forms the autonomic nervous system (ANS) at the interface of the central nervous system and the body. In this manuscript, we will focus on the VN distribution at the level of the digestive tract, as well as its anti-inflammatory properties, which can be targeted by vagus nerve stimulation (VNS) in the domain of inflammatory bowel diseases (IBD) and other inflammatory disorders.

The VN is a mixed nerve containing predominantly 80% of afferent fibers vehiculating visceral, somatic and taste sensations and 20% of efferent fibers controlling gastrointestinal motility and secretion. In particular, the VN is involved in the “conditioned reflex” described by the famous Russian researcher Ivan Petrovich Pavlov who won the Nobel Prize for Physiology or Medicine in 1904. The VN typically transmits information to the brain regarding luminal osmolarity, carbohydrate levels, mechanical distortion of the mucosa, and the presence of cytostatic drugs and bacterial products, whereas sympathetic afferents normally transmit visceral pain. Vagal efferents originate in the dorsal motor nucleus of the VN (DMNV), in the medulla, and innervate the digestive tract from the esophagus to the splenic flexure while the rest of the gut (left colon and rectum) is innervated...
by the sacral (S2-S4) parasympathetic nucleus (Netter, Colacino 1989). However, according to J. Delmas and G. Laux (1933), the VN innervates all the digestive tract in humans. In rats, S. M. Altschuler et al. (1993) reported that the VN innervates all of the digestive tract except the rectum. Vagal efferent fibers do not reach the intestinal lamina propria directly (Berthoud et al. 1991) but synapse onto enteric neurons that innervate the lamina propria where they release acetylcholine (ACh) acting on nicotinic or muscarinic receptors. Vagal afferent fibers originate from the mucosa to the muscle layers of the digestive tract. Their sensory afferent cell bodies are located in nodose ganglia and relay information to the nucleus tractus solitarii (NTS) (Cechetto 1987) and the area postrema, in close relation with the DMNV to form the dorsal vagal complex. Visceral information is then transmitted to areas of the forebrain such as the hypothalamus, amygdala, and cortex, via a relay through the parabrachial nucleus, hypothalamic-pituitary-adrenal (HPA) axis, thalamus before final visceral afferent inputs in the insular cortex, the anterior cingulate and prefrontal cortices corresponding to the central autonomic network (CAN) (Benarroch 1993). The CAN, in turn, is able to modulate the ANS especially through projections of (i) the paraventricular nucleus of the hypothalamus (PVH) to the DMNV and preganglionic neurons of the sympathetic nervous system at the spinal cord level; (ii) the amygdala to the DMN; (iii) the Barrington nucleus to the sacral parasympathetic nucleus; (iv) the A5 noradrenergic and C1 adrenergic groups to spinal preganglionic sympathetic neurons (Ricardo, Koh 1978). This complex is involved in the autonomic, endocrine and limbic responses of the “inner medium”. The VN is a major component of interoceptive pathways and is considered the sixth sense of the body (Zagon 2001). Interoception, the sense of the body’s internal physiological state, is regarded as the basis for motivational and emotional feelings and to the core neurobiological representation of self. Interoceptive mechanisms appear central to somatic disorders of brain-body interactions, including functional digestive disorders, such as irritable bowel syndrome (IBS), but also IBD (Bonaz et al. 2020; Fournier et al. 2020).

**Anti-inflammatory properties of the vagus nerve**

The VN has anti-inflammatory properties both through its afferent and efferent fibers (Fig. 1). Indeed, the release of peripheral pro-inflammatory cytokines, such as interleukin (IL)-1, IL-6, and tumour necrosis factor (TNF), by macrophages and other immune cells, following peripheral (iv or ip) injection of lipopolysaccharide (LPS), as a septic shock model (Werner et al. 2003), activates vagal afferents, through IL-1β receptors of the paraganglia. Then, the information is conveyed to the NTS, at the level of the medulla, where neurons projecting the information to the PVH, around corticotrophin-releasing-factor (CRF)-containing neurons, are activated. These CRF neurons will then induce the release of adrenocorticotropic hormone by the pituitary, finally stimulating the release of glucocorticoids by the adrenal glands to dampen peripheral inflammation, i.e. the HPA axis. Therefore, the VN is a major component of the neuro-endocrine immune axis. This anti-inflammatory pathway is disrupted by vagotomy, which worsens experimental colitis (Ghia et al. 2006). Circulating pro-inflammatory cytokines are also able to target circumventricular organs, located outside the blood-brain barrier, that stimulate neighbouring neurons, which then stimulate the HPA axis (Buller 2001).
In 2000, Tracey's group described the parasympathetic cholinergic anti-inflammatory pathway (CAP) involving the vago-vagal reflex where vagal afferent fibers activate vagal efferent fibers (Fig. 1). Indeed, peripheral iv injection of LPS inducing a septic shock in rats was prevented by VN stimulation (VNS) of the distal end cut VN, thus stimulating vagal efferent fibers (Borovikova et al. 2000). This effect was mediated by the release of ACh by the VN inhibiting the release of pro-inflammatory cytokines such as TNF by activated macrophages. Besides TNF, other pro-inflammatory cytokines such as IL6 and IL1β were significantly decreased by VNS, but not the anti-inflammatory cytokine IL-10. This anti-inflammatory effect was mediated by linking ACh with α-7-nicotinic ACh receptors (α7nAChR) of macrophages and was suppressed in α7nAChR knockout animals (Wang et al. 2003). The intra-cellular signaling of α7nAChRs inhibits transactivational activity of the transcription factor NF-kB p65 (Wang et al. 2004) and activates Jak2 and STAT3 signalling (de Jonge et al. 2005). However, the VN does not interact directly with resident macrophages in the gut but interacts with nNOS, VIP, and ChAT enteric neurons located within the gut muscularis and with their nerve endings close to resident macrophages (Cailotto et al. 2014).

Tracey's group also described a vago-sympathetic anti-inflammatory pathway through which the VN activates the splenic sympathetic nerve (Rosas-Ballina et al. 2008) (Fig. 1). Indeed, the release of norepinephrine by this splenic nerve binds to the β2 adrenergic receptor of splenic lymphocytes, which release ACh, which binds to α7nAChRs of splenic macrophages finally inhibiting the release of TNF by the spleen (Olofsson et al. 2012). This is the non-neuronal cholinergic pathway. Thus, the VN has an anti-TNF effect either at the peripheral macrophages level or at the spleen level. However, for some authors, the efferent pathway of the CAP is not the VN but the sympathetic nervous system. For example, D. Martelli et al. (2014) proposed a non-neural link from the VN to the spleen where α7nAChRs are located on the peripheral terminals of the splenic sympathetic nerves. When stimulated by ACh from incoming T-cells, these terminals release norepinephrine, which then acts on β adrenergic receptors on splenic macrophages to suppress their release of TNF. Another anatomical pathway could be the activation of the celiac ganglion, at the origin of the innervation of the spleen, by brain nuclei that are parts of the CAN and that generate patterns of autonomic responses via projections to preganglionic sympathetic neurons in the spinal cord, such as the C1 adrenergic group (Abe et al. 2017). Indeed, five cell groups in the brain regulate the entire sympathetic outflow (Strack et al. 1989): the PVN, the A5 noradrenergic cell group, the caudal raphe region, the rostral ventrolateral medulla (C1 adrenergic group), and the ventromedial medulla. The activation of the afferent arm (i.e. vagal afferents) of the inflammatory reflex could activate the CAN, through projections from the NTS, to modulate the sympathetic nervous system through these five cell groups. In this case, the VN would induce an indirect anti-inflammatory reflex by activating the sympathetic nervous system.

Thus, the anti-inflammatory pathways involving the VN are rather complex and it clearly appears that both vagal afferents and efferents are of interest in this respect. Targeting this anti-inflammatory pathway is of interest in various pathologic conditions, especially in inflammatory disorders where cytokines are involved in their pathogenicity such as IBD, represented by Crohn's disease (CD) and ulcerative colitis, and rheumatoid arthritis (RA). Presently, in the domain of IBD and RA, biological therapies, such as anti-TNF, have revolutionized the care of these patients. However, these treatments have complex side effects (Click, Regueiro 2019) resulting in 30–50% of non-adherence (Chan et al. 2017); and patients show a growing interest in complementary medicines (Torres et al. 2019). These biological therapies act downstream of macrophages and other immune cells while other therapeutic approaches can act upstream (Fig. 1). For example, (i) α7nAChRs agonists (GTS-21, AR-R17779) or nicotine have been used in inflammatory models of post-operative ileus, pancreatitis; (ii) activation of the central cholinergic pathway with CNI 1493 or Galantamine in a model of carrageenan-induced paw edema and endotoxemia improve inflammation; (iii) nutritional therapy with fat nutrition induces the release of cholecystokinin which activates vagal afferents which blunted hemorrhagic shock-induced TNF and IL-6 release; (iv) complementary therapies such as hypnosis, meditation, tai chi, acupuncture as well as physical activity stimulate the VN; and, finally, (v) VNS is of interest in activating the CAP; we will now focus on this non-drug therapy approach (see a review by Bonaz et al. 2017; 2019).

**Vagus nerve stimulation in epilepsy**

VNS was approved by the FDA for the treatment of drug-resistant epilepsy in 1997 (Bonaz et al. 2013). Today, ~100,000 patients have been implanted for epilepsy. About 50% of patients have a significant reduction in seizure frequency, with ~12% experiencing a 90% decrease in seizures.
Vagus nerve stimulation in inflammatory bowel disease

Based on the theory of the CAP, we first performed VNS in an experimental model of TNBS (2,4,6-trinitrobenzenesulfonic acid)-induced colitis (Th1-induced inflammation) in rats, resembling CD (Meregnani et al. 2011). VNS was performed in freely moving animals chronically stimulated 3 h per day for 5 days, starting 1 h before colitis, with the following stimulation parameters: 1 mA, 5 Hz, pulse width of 500 μs; 10 s ON, 90 s OFF; continuous cycle with an external stimulator. Control rats were implanted according to the same procedure but were not stimulated. VNS reduced body weight loss in rats with colitis and improved inflammatory markers above the colonic lesions as observed by histological score and myeloperoxidase quantification. This anti-inflammatory effect was also demonstrated by the improvement of a multivariate index of colitis (Meregnani et al. 2011).

In a translational bench-to-bedside approach, we performed a pilot study of VNS in patients with active CD (ClinicalTrials.gov Identifier: NCT01569503) where VNS was regarded as an alternative to anti-TNF treatment. This study received a grant from the INSERM-DGOS. Nine patients have been implanted according to the protocol and with the device (neurostimulator and electrode) for epilepsy, using the following parameters: intensity, 0.5–1.5 mA; frequency, 10 Hz; pulse width, 500 μs; stimulation on-time: 30 s, off-time: 5 min. The first patient was implanted in April 2012 and the last patient in March 2016. Patients were selected according to a clinical activity index (CDAI: Crohn’s disease activity index, 220 < CDAI < 450), biological parameters (C-reactive protein, CRP > 5 mg/L; fecal calprotectin > 100μg/g), endoscopic parameters (CDEIS: Crohn’s disease endoscopic index of severity; CDEIS ≥ 7) with the aim to induce clinical, biological, and endoscopic remission. It was a 12-month follow-up study. We published, for the first time, the 6-month follow-up of the seven first implanted patients where 5/7 patients had responded to VNS with clinical, biological and endoscopic improvement/healing, with a restored vagal tone (Bonaz et al. 2016). Two patients were withdrawn from the study after 3 months of VNS, due to worsening of their disease. One patient underwent surgery (ileocecal resection) and then was treated with a combination therapy (Infliximab + azathioprine) and the other patient received the same combination therapy. These two patients are presently in remission while the VNS device was left in place with the intensity turned down. Among the other 5 patients, only one patient was treated with an immunosuppressant (azathioprine). These preliminary results showed that VNS was feasible and could be a promising tool in the treatment of active CD. The procedure was safe with the usual side effects described above. No device was removed during the study period and later on. Recently, we published the 12-month follow-up data on 9 patients (Sinniger et al. 2020). After 1 year of VNS, five patients were in clinical remission and six in endoscopic remission. CRP
and fecal calprotectin decreased in six and five patients, respectively. Seven patients restored their vagal tone and decreased their digestive pain score. The patients’ cytokinergic profile evolved toward a more “healthy profile”: IL-6, -23, -12, TNF and transforming growth factor β1 were the most impacted cytokines. Correlations were observed between CRP and TNF, and some gut mucosa metabolites such as taurine, lactate, alanine and beta-hydroxybutyrate. VNS was well tolerated, as reported in the 6-month follow-up study.

These results are in agreement with the preliminary results of another study published in an abstract form (D’Haens et al. 2018), which observed clinical, biomarker, and endoscopic improvement for half of the 16 CD patients who received either VNS monotherapy (biologics refractory patients) or VNS adjunctive therapy for 4 months. We also showed that, in addition to the activation of vagal efferent fibers that regulate the ANS, our data suggest that chronic VNS has a regulatory effect via afferent vagal fibers on anxiodepressive symptomatology associated with CD, which could be directly highlighted by the modulation of EEG alpha power known to be associated with depressed states (Kibleur et al. 2018).

VNS in other inflammatory conditions

RA is also a chronic inflammatory condition where VNS is of interest. Experimental data have shown that knockdown of the α7nAChR in RA fibroblast-like synoviocytes increased the production of mediators of inflammation, and degradation and activation of α7nAChRs in an animal model of RA resulted in reduced arthritis activity (Koopman et al. 2014). Accordingly, stimulation of the CAP by VNS improved an experimental model of arthritis while aggravation of arthritis activity was observed after unilateral cervical vagotomy, as well as in α7nAChR-knockout mice. In 2016, F. A. Koopman et al. (2016) showed — using an implantable device from SetPoint Medical Corporation (Valencia, CA) — that VNS (up to four times daily) significantly inhibited TNF production for up to 84 days in patients with RA. In addition, RA disease severity, measured by standardized clinical composite scores, improved significantly; thus suggesting that it is possible to use neuroimmune modulating device in the therapy of RA with potential implications in other auto-immune and auto-inflammatory diseases. Postoperative ileus (POI) is characterized by a delay of gastric emptying and prolonged intestinal transit after surgery (Stakenborg et al. 2017a). Postoperative ileus can prolong hospitalization and increase healthcare costs. A peripheral pathway, involving the CAP, has shown the release of TNF by resident macrophages in the muscularis propria after abdominal surgery (de Jonge et al. 2003). VNS improves postoperative ileus both in experimental conditions and in humans (Stakenborg et al. 2017b).

Non-invasive vagus nerve stimulation

The data that we have reported above were based on invasive VNS that requires a surgical implantation of an electrode and a neurostimulator. However, the development of non-invasive VNS techniques is of interest, such as transcutoaneous auricular VNS (ta-VNS). This technique is based on the innervation pattern of the lateral surface auricle of the cymba conchae, cavity of the concha, and tragus which are innervated by the auricular (afferent) branch of VN, respectively at 100%, 45%, and 45% (Peuker, Filler 2002). Thus, stimulation of this anatomical part of the auricle will activate vagal afferents targeting the NTS and then activating vagal efferent through the inflammatory reflex described by Tracey’s group. Afferent pathway of the VN and regions activated by taVNS (fMRI studies) are the same (Badran et al. 2018). The Cerbomed Nemos device (Erlangen, Germany) is an external device that provides ta-VNS by using a dedicated intra-auricular electrode (like an earphone) which stimulates the auricular branch of the VN (Stefan et al. 2012). This device received the European clearance (CE mark) in 2010 for epilepsy treatment and is currently available in Germany, Austria, Switzerland, and Italy. Another ta-VNS device is marketed by Schwa Medico (Rouffach, France) with an electrode stimulating the cymba conchae and the cavity of the concha and connected to a neurostimulator (Urostim 2); this device is recommended for IBS (Mion et al. 2020), IBD, RA, migraine, pelvic pain and fibromyalgia treatment. The Elecrocore LLC Gammacore device (Basking Ridge, NJ, USA) is a transcutoaneous cervical VN stimulator that uses proprietary electrical signals to treat primary headache (Mwamburi et al. 2020). These non-invasive devices are well tolerated with no major side effects, and could be used for the treatment of various inflammatory disorders and others, because the VN has anti-inflammatory but also anti-nociceptive and anti-depressive properties.

Conclusion

The anti-inflammatory properties of the VN, both through its afferents (activation of the HPA axis) and efferents (activation of the CAP) can be used to treat various pathological conditions such as chronic inflammatory diseases. In particular,
as a key element of the ANS in the brain-gut interactions in IBD (Bonaz et al. 2013), the VN is a therapeutic target in IBD. In addition, the VN is also at the interface of the microbiota-gut-brain axis (Bonaz et al. 2018) and based on the implication of dysbiosis in various pathological conditions, one can assume that VNS is also able to target this dysbiosis. VNS is able to restore vagal tone. Indeed, we have reported an abnormal ANS in IBD patients (Pellissier et al. 2010) negatively correlated with TNFα levels (Pellissier et al. 2014). VNS, by restoring the ANS balance in such patients through the activation of the VN, is a novel therapeutic treatment. Finally, VNS is devoid of the adverse events of biological therapies such as anti-TNF drugs (infections, dermatological complications, lymphoma, immunisation) which are still a gold standard treatment in IBD but are feared by patients, thus explaining their non-adherence to treatment and their search for a non-drug therapy. Finally, the economic impact of VNS is of interest since, for example, the median cost of 1-year anti-TNF therapy raises up to $40,000 for CD patients (Targownik et al. 2019). The use of neuromodulation by bioelectronics devices as a treatment is an emerging field in the domain of bioelectronic medicine. It could be an alternative non-drug therapy to conventional treatment or could be combined with such treatments, but further investigation in large longitudinal cohorts of patients is required. Finally, the optimal neurostimulation parameters to better treat common conditions and diseases that involve immune regulation to achieve significant cytokine changes need to be determined (Bonaz 2020).

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